

# University of Virginia Institutional Biosafety Committee Guidelines for Biosafety Level Assignment of Adeno-Associated Viral Vectors (AAV)

Adopted by the IBC November 12, 2013. Revised: 2/2/2016; 2/7/2017; 06/25/2024

**Background:** Adeno-associated viruses are members of the parvovirus family. They are non-enveloped, single-stranded DNA viruses that can only replicate in the presence of a helper virus such as adenovirus (Ad), herpes simplex virus, human papillomavirus or vaccinia for productive infection. Recombinant AAV-vectors of various serotypes which differ in their cell tropism are widely used for gene delivery in biomedical research as their characteristics include: the ability to infect a broad range of dividing and non-dividing cells; ability to be produced in high titers; long-term, stable expression and different serotypes specifically bind to different target cell populations.

*Potential AAV Health Hazards:* While AAV is generally considered non-pathogenic; there are several reports describing AAV infection in association with adverse reproductive outcomes, including spontaneous abortion and spontaneous preterm birth<sup>1, 2, 3, 6</sup>. In males, integrated AAV DNA has been detected in testis tissue sample and the presence of AAV DNA in semen samples has been reported and suggested to be associated with male infertility<sup>3, 5</sup>. Further, while wild type AAV integrates preferentially into human chromosome 19 and remains latent until a helper virus supplies the necessary genes for replication, recombinant AAV vectors lose site specific integration, thereby raising the theoretical concern of insertional mutagenesis. While 90% of humans are seropositive for AAV, genomic analysis of tissues demonstrated AAV sequences are found in a variety of tissues with a hybrid of two serotype of AAV commonly found integrated in human liver and bone marrow<sup>4</sup>. Recent findings also provide evidence that chromosomal insertions of AAV serotype 2 increases the expression of proto-oncogenes in human hepatocellular carcinoma<sup>7</sup>.

*Residual Human Cell Contaminants:* AAV vectors are typically propagated in HEK 293 cells either in the presence of a helper virus or by co-transfection of helper plasmids. HEK 293 are a commercially available human embryonic kidney continuous cell line. [The UVA IBC Policy on the Use of Human Derived Material](#) requires that all cell and organ cultures of human origin, including well established cell lines be handled in accordance with the [OSHA Bloodborne Pathogens Standard](#) under BSL2 containment. Unless viral purification and quality control measures for AAV are effectively applied, it is possible that any contaminants that may have been present in the original human cell culture will also be contained in the resulting supernatant.

[The NIH Guidelines for Research Involving Recombinant or Synthetic DNA Molecules](#) identify all AAV serotypes and recombinant or synthetic AAV constructs as risk group 1 (RG1) agents so long as the transgene does not encode either a potentially tumorigenic gene product (e.g. oncogene) or a toxin molecule and is produced in the absence of a helper virus.

### **Use of AAV at BSL1/ABSL1 Containment:**

The IBC will consider requests for designating AAV experiments at BSL1/ABSL1 containment if the following information is provided:

- 1) The nature of transgene expression including gene ID and function.
- 2) Use of adenovirus or any other helper virus of human origin in AAV vector production
- 3) Identification of the cell line used to propagate the AAV vector
- 4) Core facility/company/laboratory providing the AAV vector
- 5) A description of purification procedures (e.g. column chromatography, etc.) and/or documentation from the source facility that describes purification procedures and/or purity
- 6) If applicable, a description of procedures to ensure AAV vectors used under BSL2/ABSL2 containment are kept separate from those approved at BSL1/ABSL1

**Use of AAV at BSL2/ABSL2: Proposed experiments involving use of AAV that do not satisfy the above criteria for BSL1/ABSL1 containment must be conducted at BSL2/ABSL2 containment.**

*We acknowledge and express our appreciation to the University of Iowa Environmental Health and Safety Department and the University of Pittsburgh for use of their AAV Guidance documents in the development of this policy.*

### **References**

1. Arechavaleta-Velasco F, Gomez L, Ma Y, Zhao J, McGrath CM, Sammel MD, Nelson DB, Parry S. Adverse reproductive outcomes in urban women with adeno-associated virus-2 infections in early pregnancy. *Hum Reprod* 2008; 23:29-36.
2. Burguete, T. *et al.* Evidence for infection of the human embryo with adeno-associated virus in pregnancy. *Human Reproduction*. 1999; 14:2396-2401.
3. Chung HK, *et al.* Detection of Adeno-associated Virus from Semen Suffering with Male Factor Infertility and Having Their Conception Partners with Recurrent Miscarriages. *J Bacteriology and Virology* 2012; 42:339-345.
4. Gao G, Vandenberghe LH, Alvira MR, Lu Y, Calcedo R, Zhou X, and Wilson JM. Clades of Adeno-Associated Viruses Are Widely Disseminated in Human Tissues. *J. Virology* 2004; 78: 6381–6388.
5. Mehrle S, Rohde V, Schlehofer JR. Evidence of chromosomal integration of AAV DNA in human testis tissue. *Virus Genes* 2004; 28:61-69.
6. Tobiasch E, Rabreau M, Geletneky K, Larue-Charlus S, Severin F, Becker N, Schlehofer JR Detection of adeno-associated virus DNA in human genital tissue and in material from spontaneous abortion. *J Med Virol* 1994; 44:215-222.
7. Nault, JC *et al.* Recurrent AAV2-related insertional mutagenesis in human hepatocellular carcinomas. *Nature Genetics* 2015; 47:1187-1193.